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- 1 Author to whom correspondence should be sent.
- 2 Incumbent of the Graham and Rhona Beck Career Development Chair, Isotope Department, The Weizmann Institute of Science, 76 100 Rehovot, Israel.
- 3 Lowenstam, H. A., *Chem. Geol.* 9 (1972) 153.
- 4 Lowenstam, H. A., *Science* 213 (1981) 1126.
- 5 Lowenstam, H. A., and Margulis, L., *BioSystems* 12 (1980) 27.
- 6 Lowenstam, H. A., and Weiner, S., in: *Biomining and Biological Metal Accumulation*, p. 191. Eds P. Westbroek and E. W. de Jong. D. Reidel, Amsterdam 1983.
- 7 Gibson, R., *Nemertean. Hutchinson and Co.*, 1972.
- 8 Stricker, S. A., and Cloney, R. A., *Zoomorphology* 97 (1981) 205.
- 9 Stricker, S. A., and Cloney, R. A., *Biol. Bull.* 162 (1982) 387.
- 10 Stricker, S. A., *J. Morph.* 175 (1983) 182.
- 11 Stricker, S. A., *J. Morph.* 179 (1984) 119.

- 12 Wourms, J. P., *Am. Zool.* 16 (1976) 213.
- 13 Stricker, S. A., Cavey, M. J., and Cloney, R. A., *Trans. Am. microsc. Soc.* 104 (1985) 232.
- 14 Termine, J. D., and Posner, A. S., *Science* 153 (1966) 1523.
- 15 Featherstone, J. D. B., Pearson, S., and LeGeros, R. Z., *Caries Res.* 18 (1984) 63.
- 16 Lowenstam, H. A., and Weiner, S., *Science* 227 (1985) 51.
- 17 Blumenthal, N. C., Posner, A. S., Silverman, L. D., and Rosenberg, L. C., *Calcif. Tiss. Int.* 27 (1979) 75.
- 18 Becker, G. L., Chen, C.-H., Greenawalt, J. W., and Lehninger, A. L., *J. Cell Biol.* 61 (1974) 316.
- 19 Watabe, N., *Science* 124 (1956) 630.
- 20 McConnell, D., *Bull. geol. Soc. Am.* 74 (1963) 363.
- 21 Lowenstam, H. A., *Science* 156 (1967) 1373.
- 22 Neff, J. M., *Calcif. Tiss. Res.* 7 (1971) 191.

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## Biliverdin as an electron transfer catalyst for superoxide ion in aqueous medium

G. Galliani, D. Monti, G. Speranza and P. Manitto

*Dipartimento di Chimica Organica e Industriale dell'Università and Centro di Studio per le Sostanze Organiche Naturali del CNR, Via Venezian 21, I-20133 Milano (Italy), and Centro Studi Maria Branca, Via E. Porro 1, I-20158 Milano (Italy), 26 November 1984*

**Summary.** Stopped flow experiments gave evidence of the formation of a biliverdin-superoxide complex and/or a biliverdin radical anion by reaction of aqueous  $O_2^-$  with biliverdin. Such transient species are likely intermediates both in the bleaching of biliverdin, during exposure to the aerobic xanthine oxidase reaction, and in the reduction of ferricytochrome *c* under the same conditions.

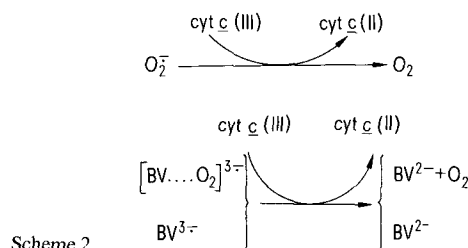
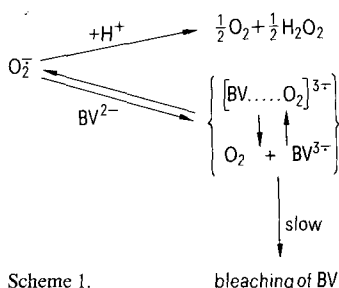
**Key words.** Biliverdin; superoxide ion; cytochrome *c*; stopped flow technique; xanthine oxidase; superoxide dismutase.

Although the bleaching of biliverdin (BV)<sup>1</sup> during exposure to the aerobic xanthine oxidase reaction has been reported by Fridovich<sup>2</sup>, the chemical processes involved remain to be clarified. Recently we found that BV (and its dimethyl ester) interacts rapidly with  $KO_2$  in DMSO, giving rise to a reversible 1:1 adduct<sup>3</sup>. This prompted us to investigate whether a similar charge-transfer complex could be the actual intermediate in the reaction of BV with enzymatically generated superoxide<sup>4</sup>. In this paper we give spectroscopic evidence of the formation of radical anions, such as the complex  $[BV \cdots O_2]^{3-}$  or  $BV^{3-}$  or both, as likely transient intermediates in the bleaching of BV by aqueous  $O_2^-$  and in the BV-catalyzed reduction of ferricytochrome *c* by the same reagent. In studying the system BV/ $KO_2$  in DMSO a thermodynamic approach (i.e. chemical equilibrium determination) was followed<sup>3</sup>. Such a method, however, appeared to be unsuitable in the case of aqueous media since the superoxide ion undergoes a rapid dismutation (to  $O_2$  and  $H_2O_2$ ) in protic solvents<sup>5</sup>. For this reason a kinetic approach, based on a stopped flow technique, was chosen. When BV<sup>6</sup>, dissolved in an oxygenated solution of xanthine, was mixed with an oxygenated solution of xanthine oxidase in stopped flow apparatus, a transient

species A could be detected showing an electronic absorption maximum at 730 nm (figs 1 and 2).

Equal volumes of two solutions containing xanthine-oxidase in phosphate buffer pH 7.6 and a mixture of xanthine and biliverdin in DMSO were mixed in a stopped flow apparatus (Nortech Laboratories Limited, England, model FPX-1; mixing time 5 ms) according to the conditions described in the captions of the figures. The transient signals were recorded by means of a Tektronix model 5115 oscilloscope and a Polaroid camera. The base line of the starting spectrum was practically restored after the decay of A (within 200 ms), indicating that the BV bleaching occurs on a completely different time scale (cf. Robertson Jr and Fridovich<sup>2</sup>). Formation of A was not observed when deoxygenated solutions were used, and the concentration of A was lowered to zero by the addition of increasing amounts of bovine erythrocyte superoxide dismutase. Thus, figure 2 can be regarded as a pre-steady state of the reaction of BV with  $O_2^-$  generated by the system xanthine/xanthine oxidase/oxygen.

It must be pointed out that the reported spectrum of the BV radical anion at basic pH ( $BV^{3-}$ )<sup>7</sup> fits the spectrum of A nearly exactly<sup>8</sup>. However, one cannot rule out that A is a charge-trans-



fer complex, viz.  $[BV \cdots O_2]^{3-}$ , like that formed by interaction of BV with  $KO_2$  in DMSO (which shows a comparable spectrum)<sup>3</sup>. We cannot even rule out that the charge-transfer complex is actually formed to give successively dioxygen and  $BV^{3-}$  through a rapid internal electron transfer (see scheme 1). Whatever the structure of A may be, it is noteworthy that its formation seems to be associated with a rate enhancement of reactions involving the superoxide ion as a one-electron reductant, as shown by the following findings.

The reduction of ferricytochrome *c* by superoxide has been largely utilized as an indirect method to detect and to measure production and decay of  $O_2^-$  in aqueous solution<sup>2,4</sup>. The addition of BV to a system containing cytochrome *c*, xanthine, xanthine oxidase and molecular oxygen resulted in a sharp increase

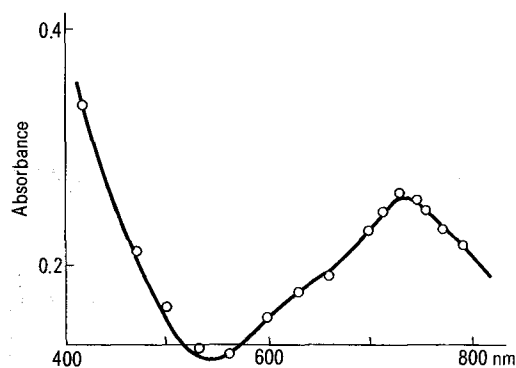


Figure 1. Difference spectrum of A. Absorbances at different wavelengths were recorded 45 ms after mixing a solution of xanthine and BV with a solution of xanthine oxidase (i.e. at the maximum concentration of A; see fig. 2). Conditions:  $T = 21^\circ\text{C}$ , phosphate buffer 0.03 M, pH 7.6,  $O_2$  at saturation; final concentrations: xanthine  $10^{-3}$  M, BV  $10^{-4}$  M, xanthine oxidase 10 mU/ml.

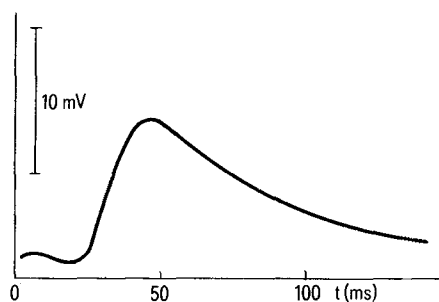


Figure 2. Oscillogram recorded at 730 nm from a stopped flow experiment. Conditions as in figure 1.

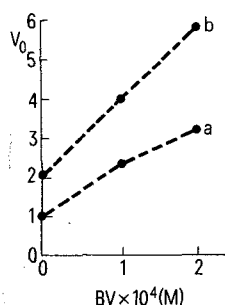


Figure 3. Initial rate of cytochrome *c* reduction (in arbitrary units) vs BV concentration. Conditions:  $T = 21^\circ\text{C}$ , phosphate buffer 0.1 M pH 7.6, xanthine  $10^{-3}$  M, xanthine oxidase 4 mU/ml,  $O_2$  at saturation, cytochrome *c*: (a)  $2 \times 10^{-5}$  M, (b)  $4 \times 10^{-5}$  M.

of the rate of cytochrome *c* reduction, as shown in figure 3<sup>9</sup>. Curves a and b in figure 3 differ when  $BV = 0$  because the reduction of cytochrome *c* by  $O_2^-$  is first order in cytochrome *c* under the experimental conditions. The rate constant for the reaction of superoxide ion with cytochrome *c* has been estimated as  $1.1 \times 10^5 - 1.1 \times 10^6 \text{ M}^{-1} \text{ s}^{-1}$ , whereas larger rate constants have been reported in the oxidation of free radicals by ferricytochrome *c*<sup>10</sup>.

The different time scale for the reduction of ferricytochrome *c* by superoxide, which is accomplished within a few minutes, and the bleaching of biliverdin (lasting more than 1 h,  $k = 7 \times 10^3 \text{ M}^{-1} \text{ s}^{-1}$ ) rules out that any radical fragment deriving from the latter process can be responsible for the increase of the rate of the former. Thus, a plausible explanation of this fact is depicted in scheme 2, pivoting on the transient species A and taking into account all the hypotheses mentioned above about the nature and the origin of A. The following assumptions can be made: a) a steady state concentration of A is reached after ca. 200 ms after starting the reaction producing superoxide; b) superoxide reacts with BV more rapidly than with ferricytochrome *c*; c) ferricytochrome *c* is reduced more rapidly by A than by superoxide.

A steady state concentration of A could also explain the slow bleaching of BV (scheme 1): if A is the BV radical anion, this is expected to disproportionate to give BV and bilirubin, which is rapidly destroyed by superoxide<sup>2</sup>. If A is a charge-transfer complex, it could be bleached via hydrogen abstraction as shown previously<sup>3</sup>. Thus figure 2 can be interpreted as follows. The upward side of the curve corresponds to the direct reaction of  $BV^{2-}$  with  $O_2^-$ , whereas the downward side is related to at least two processes: the reverse reaction of the equilibrium between  $BV^{2-}$  and  $O_2^-$ , whose driving force is given by superoxide dismutation, and the bleaching of A. After a brief pre-steady state, the steady state concentration of A is obtained, but it is so low that it cannot be estimated by its absorption. This interpretation is reinforced by the observation that, introducing another consecutive reaction of A by adding ferricytochrome *c*, the maximum concentration of A (observable as in fig. 2) is lowered.

While it is well known that the reactivity of superoxide ion can be affected by its complexation with metal ions<sup>5</sup>, its interaction with a biological molecule (biliverdin) represents, to our knowledge, the first example of a reactivity enhancement by the intermediacy of a rapidly formed radical species.

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- 1 Biliverdin plays a key role in the catabolism of heme in that it is formed in vivo as the first isolable product of the oxidative breakdown of the protoporphyrin ring: see Schmid, R., and McDonagh, A. F., in: *The Porphyrins*, vol. 6, p. 257. Ed. D. Dolphin, Academic Press, New York 1979.
- 2 Robertson, P. Jr, and Fridovich, I., *Archs Biochem. Biophys.* 213 (1982) 353.
- 3 Galliani, G., Monti, D., Speranza, G., and Manitto, P., *Tetrahedron Lett.* 25 (1984) 6037.
- 4 Fridovich, I., *J. biol. Chem.* 245 (1970) 4053.
- 5 Sawyer, D. T., and Valentine, J. S., *Acc. Chem. Res.* 14 (1981) 393.
- 6 Manitto, P., and Monti, D., *Experientia* 35 (1979) 9.
- 7 Land, E. J., Sloper, R. W., and Truscott, T. G., *Radiation Res.* 96 (1983) 450.
- 8 Both  $CO_2H$  groups of BV are likely dissociated under the reaction condition (pH 7.6<sup>2</sup>). See McDonagh, A. F., in: *The Porphyrins*, vol. 6, p. 293. Ed. D. Dolphin, Academic Press, New York 1979.
- 9 This could explain some kinetic discrepancies observed by Fridovich using cytochrome *c*, instead of superoxide dismutase, as a competitor of BV for  $O_2^-$  (see Robertson Jr and Fridovich<sup>2</sup>).
- 10 Ferguson-Miller, S., Brautigan, D. L., and Margoliash, E., in: *The Porphyrins*, vol. 7, p. 200. Ed. D. Dolphin, Academic Press, New York 1979.